
LEAF-STATE-ANALYZER LSA-2050

Manual

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Contents

1	SAFETY INSTRUCTIONS	3
1.1	<i>General Safety Instructions.....</i>	3
1.2	<i>Special Safety Instructions.....</i>	4
2	INTRODUCTION	5
3	LSA-2050: EXTENT OF DELIVERY	7
3.1	<i>LSA-2050 Description</i>	8
4	ACCESSORY	11
4.1	<i>Darkening Bags LSA-2050/DB</i>	11
5	OPERATION	13
5.1	<i>Quick Start.....</i>	14
5.2	<i>Detailed Instructions</i>	14
5.2.1	<i>Main Windows.....</i>	14
5.2.2	<i>Main Menu.....</i>	17
5.2.2.1	<i>Settings</i>	17
5.2.2.2	<i>Calibration</i>	20
5.2.2.3	<i>Create New File.....</i>	21
5.2.2.4	<i>Memory</i>	21
5.2.2.5	<i>Device Info.....</i>	22
5.3	<i>LSA-2050 Software.....</i>	22
5.3.1	<i>Installing the Software.....</i>	22
5.3.2	<i>Connecting the LSA-2050 to Computer.....</i>	23
5.3.3	<i>Adjusting Time</i>	24
5.3.4	<i>Downloading Data</i>	24
5.3.5	<i>Excel File.....</i>	26
6	COMMENTS ON RESULTS	33
6.1	<i>Fv/Fm.....</i>	33
6.2	<i>Screening</i>	34
6.3	<i>Chlorophyll Concentration</i>	35
6.4	<i>Nitrogen Balance Index (NBI),.....</i>	36
7	DOCUMENTATION	39
7.1	<i>Chlorophyll Calibration</i>	39
7.1.1	<i>Plant Species.....</i>	39

Contents

7.1.2	<i>Growth Conditions</i>	<i>40</i>
7.1.3	<i>Chl. Measurements.....</i>	<i>40</i>
7.1.4	<i>Correlations.....</i>	<i>42</i>
7.2	<i>Chlorophyll and Small Leaves</i>	<i>45</i>
7.3	<i>Screening</i>	<i>45</i>
7.3.1	<i>Mesophyll Reference Factor $RF(\lambda)$.....</i>	<i>47</i>
7.3.2	<i>Calibration and Measurement</i>	<i>50</i>
7.4	<i>Geodata.....</i>	<i>51</i>
8	SPECIFICATIONS	57
8.1	<i>LEAF-STATE-ANALYZER LSA-2050.....</i>	<i>57</i>
8.1.1	<i>General Design.....</i>	<i>57</i>
8.1.2	<i>Measuring Modules.....</i>	<i>58</i>
8.1.3	<i>Geospatial Data.....</i>	<i>58</i>
8.1.4	<i>Walz Calibration Standards LSA-2050/STD.....</i>	<i>59</i>
8.2	<i>Carrying Case LSA-2050/T</i>	<i>59</i>
8.3	<i>Battery Charger</i>	<i>59</i>
8.4	<i>Accessory</i>	<i>59</i>
8.4.1	<i>Darkening Bags LSA-2050/DB</i>	<i>59</i>
9	GUARANTEE	61
9.1	<i>Manufacturer's Guarantee</i>	<i>61</i>
9.2	<i>Conditions.....</i>	<i>61</i>
9.3	<i>Instructions</i>	<i>62</i>
9.4	<i>Applicable law</i>	<i>63</i>
10	INDEX.....	65
11	LIST OF FIGURES	67
12	LIST OF TABLES.....	69

1 Safety Instructions

1.1 General Safety Instructions

- Read and follow safety and operating instructions prior to operation of the device. Pay attention to all safety warnings.
- The LEAF-STATE-ANALYZER 2050 (LSA-2050) is designed for outdoor use. However, avoid exposing the LSA-2050 to dust, sand, and dirt as much as possible.
- Avoid taking measurements in precipitation. The device is not waterproof.
- Ensure that neither liquids nor foreign bodies get inside the LSA-2050.
- Do not put the LSA-2050 near sources of heat.
- The LSA-2050 must not be dropped. Open battery compartment only in dry and clean environment.
- Use only type AAA (Micro) batteries.
- Keep the USB-C socket clean.
- Do not open the housing of the LSA-2050. There are no serviceable parts inside. The device may only be repaired by the manufacturer.

1.2 Special Safety Instructions

- The optical components of the upper and lower leaf clamps are covered with a fragile quartz disc. Avoid exerting any force on these quartz discs.
- Do not measure moist or wet samples. Wipe samples dry before measuring.
- When operating with the lower part removed, cover open electric contacts with the special electronics lid (see Fig. 5, page 10).
- The LSA-2050 is a highly sensitive measuring system which should be only used for research purposes, as specified in this manual. Follow the instructions of this manual to avoid potential harm to the user and damage to the instrument.
- The LSA-2050 can emit very strong light! To avoid harm to your eyes, never look directly at the LEDs of the sample clip.

2 Introduction

The LEAF-STATE-ANALYZER LSA-2050 is a handheld device for non-invasive leaf analysis. The measuring device employs four different approaches to probe the plant health status: (1) the extent of protection from ultraviolet and strong visible radiation, (2) the chlorophyll concentration, and (3) the maximum photochemical quantum yield of photosystem II, F_V/F_M , and (4) the state of nitrogen supply. In summary, the LEAF-STATE-ANALYZER LSA-2050 provides a picture of stress effects and a plant's ability to cope with stress.

General Features

The LEAF-STATE-ANALYZER LSA-2050 evaluates radiation screening by measuring the intensity of chlorophyll fluorescence excited by constant radiation of specific wavelengths. The excitation intensity used is low. This is to avoid variations in the fluorescence quantum yield caused by energy quenching processes.

The four different excitation wavebands employed can be related to radiation absorption of four pigment groups: UV-B and UV-A to hydroxycinnamic acids and flavonoids, respectively [1], blue to carotenoids [2], and green to anthocyanins [3]. Absorbance values indicating relative flavonoid and anthocyanin concentration are provided.

Chlorophyll concentration is measured by the Cerovic method [4]. The method excels by high response even at high chlorophyll concentrations.

The nitrogen balance index (NBI) is calculated according to another paper by Cerovic and colleagues [5] as the ratio of chlorophyll divided by epidermal flavonoids.

Photosystem II is analyzed by the well-proven PAM fluorescence/saturation pulse method [6].

With each measurement, GPS data, leaf orientation, and the direction of sun radiation are logged.

[1] Bilger W, Veit M, Schreiber L, Schreiber U (1997) Measurement of leaf epidermal transmission of UV radiation by chlorophyll fluorescence. *Physiol Plant* 101: 754–763. <https://doi.org/10.1111/j.1399-3054.1997.tb01060.x>

[2] Nichelmann L, Schulze M, Herppich WB, Bilger W (2016) A simple indicator for non-destructive estimation of the violaxanthin cycle pigment content in leaves. *Photosynth Res* 128: 183-193. <https://doi.org/10.1007/s11120-016-0218-1>

[3] Cerovic ZG, Moise N, Agati G, Latouche G, Ben Ghazlen N, Meyer S (2008) New portable optical sensors for the assessment of winegrape phenolic maturity based on berry fluorescence. *J Food Compos Anal* 21: 650-654. <https://doi.org/10.1016/j.jfca.2008.03.012>

[4] Cerovic ZG, Masdoumier G, Ben Ghazlen N, Latouche G (2012) A new optical leaf-clip meter for simultaneous non-destructive assessment of leaf chlorophyll and epidermal flavonoids. *Physiol Plant* 146: 251–260. <https://doi.org/10.1111/j.1399-3054.2012.01639.x>

[5] Cerovic ZG, Ben Ghazlen N, Milhade C, Obert M, Debuissou S, Le Moigne M (2015) Nondestructive Diagnostic test for nitrogen nutrition of grapevine (*Vitis vinifera* L.) based on Dualex leaf-clip measurements in the field. *J Agric Food Chem* 63 (14): 3669–3680

[6] Schreiber U, Schliwa U, Bilger W (1986) Continuous recording of photochemical and non-photochemical chlorophyll fluorescence quenching with a new type of modulation fluorometer. *Photosynth Res* 10: 51-62. <https://doi.org/10.1007/bf00024185>

3 LSA-2050: Extent of Delivery

Item	Order Number
LEAF-STATE-ANALYZER	LSA-2050
Four-position battery charger	000190101116
2 x 4 eneloop AAA rechargeable batteries	000160101989
Protection plate (covers electrical contacts when operated without the lower clip jaw)	000247001714
Walz LSA-2050 set of standards	LSA-2050/STD
Allen wrench 2.5 mm	000160201201
USB C to USB A Cable	000130606258
Carrying Case	LSA-2050/T
Software LSA-2050 on USB flash drive	
LSA-2050 Manual	

Fluorescence Standard



Offset Standard



Light Scattering Standard



Calibrate Offset:
Light Scattering
Standard
plus
Offset Standard



Fig. 1: Fluorescence and Offset Standards

Calibration with the fluorescence standard is required to measure radiation screening (see Section 7.3, page 45). The signals in the presence of a sandwich of the light scattering standard (top) and the black standard (bottom) are considered as instrument offset which are subtracted from the corresponding sample signals. The light scattering standard is used to calibrate the 100% state (absence of chlorophyll) under light scattering conditions similar to a leaf.

To correctly measure chlorophyll contents, the entire optical window of the FR, NIR Emitter must be covered by the sample. In case of small samples, the optical window can be narrowed by a ring which reduces the optical window to a spot with 6 mm diameter (Fig. 2).



Fig. 2: 6 mm Diameter Spot for Small Leaves

The spot must be placed on the optical window of the FR, NIR emitter (see Fig. 3 for a device description). See also Fig. 15, page 45.

3.1 LSA-2050 Description

The tip of the LSA-2050 LEAF-STATE-ANALYZER is formed by a bipartite measuring head (Fig. 3). The heads form a clip in which a sample or standard (Fig. 1) is placed. The clip can be opened by pressing the lever on the bottom side of the device (Fig. 5).

The upper “Emitter Detector Head” holds the photodiode detector, and 5 LEDs emitting in the UV or the visible range (Fig. 4). A far red and a near infrared LED is situated in the lower “FR, NIR Emitter Head”. The emission by these LEDs has to pass through the sample to reach the photodiode. The sample’s chlorophyll concentration is derived from the beam attenuation by the sample.

The emitter detector head excites and detects fluorescence from the same side. The sample properties investigated are the

state of photosystem II and screening of ultraviolet and visible radiation.

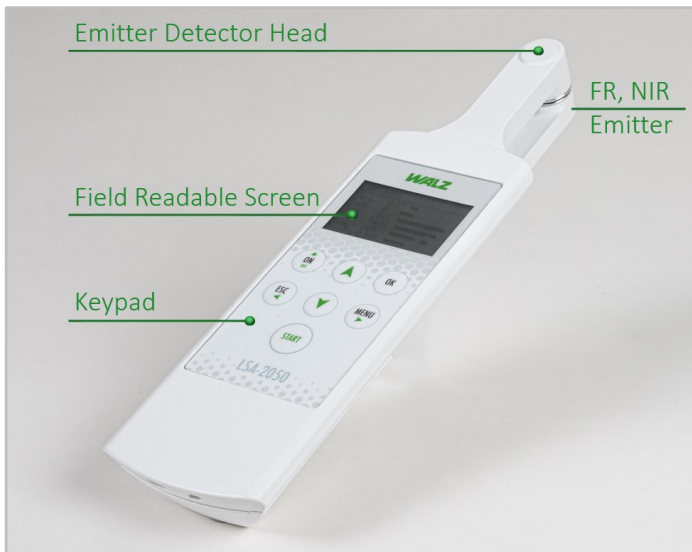


Fig. 3: LSA-2050 Overview

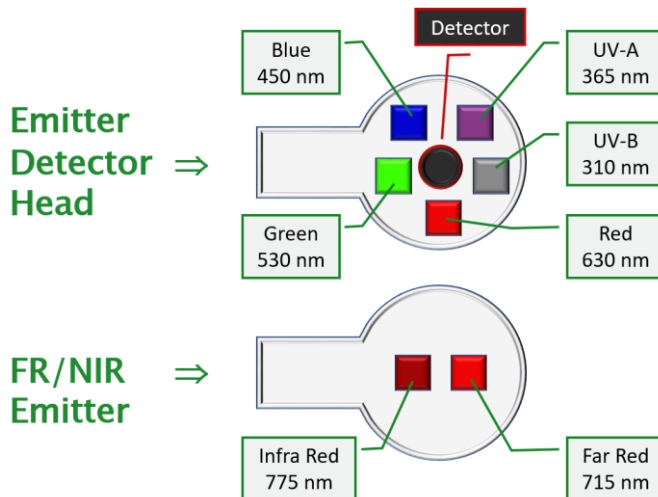


Fig. 4: LSA-2050 Measuring Head

To examine bulky samples (e.g. fruits), the FR, NIR Emitter can be removed (Fig. 5). It is obvious that chlorophyll concentrations cannot be measured with this configuration. However, it is possible to use the F_0 fluorescence as a measure for changes in the concentration of chlorophyll.

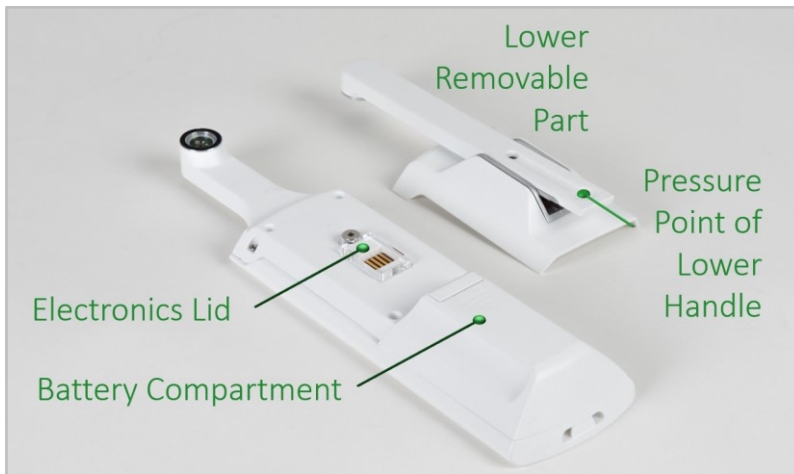


Fig. 5: LSA-2050 Modified for Bulky Samples

4 Accessory

4.1 Darkening Bags LSA-2050/DB

This accessory is designed for darkening leaves in the field. Dark acclimation is prerequisite to measure the maximum photosystem II quantum yield, F_v/F_m . The bags consist of light-tight material. Chlorophyll concentration can be determined through a central hole. The bags are available in three sizes. For details see Section 8.4.1, page 59.

A**B**

Fig. 6: Darkening Bags LSA-2050/DB

A: Darkening bags are available in three sizes. The LSA-2050/DB represents a set of bags and includes three bags of each size. **B:** Each bag has a central hole. For dark acclimation, the hole is covered on both sides by flaps.

5 Operation

The LSA-2050 is controlled by 7 keys. The function of these keys is outlined in Fig. 7. The symbols on the keys and the instructions in the software make the operation of the unit self-explanatory.

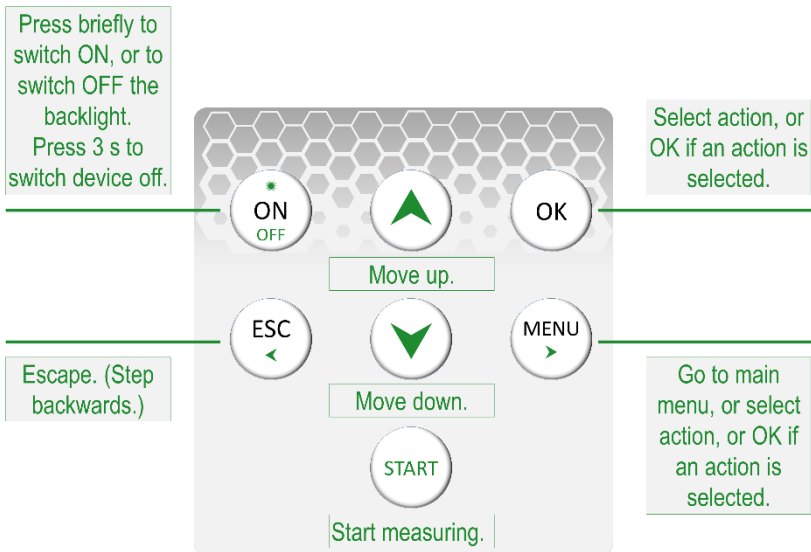


Fig. 7: Keys

The button START is only active when one of the four main windows is visible (see Section 5.2.1, page 14).

5.1 Quick Start

- Connect LSA-2050 to computer. Install and launch LSA-2050 software. Set time zone (see 5.3.3, page 24).
- On the LSA-2050, click the MENU button, select “Calibration”, and perform all four calibration procedures. Follow exactly the instructions on the screen.
- Start measuring.

Default settings apply unless changed by the user. Defaults settings are: (1) All tests active, (2) chlorophyll concentration is calculated with the calibration curve for C₃ leaves (Table 21, page 43) and it is given in nmol/cm², and (3) the mesophyll reference is from lower leaf sides of *Valeriana locusta*.

5.2 Detailed Instructions

5.2.1 Main Windows

After system start, the LSA-2050 window appears (Table 1). The window displays the apparent transmittance at the wavelengths 310, 365, 450 and 530 nm (Q_{310} , Q_{365} , Q_{450} , and Q_{530} , respectively). The window further displays the chlorophyll concentration in the unit selected in the menu “Chl. Settings” (Table 9, page 18), and the maximum photochemical yield of photo-system II, F_v/F_m . See Section 6 (page 33) for comments on this data.

The device status is indicated by symbols in the top right corner of the LSA-2050 window. The same information is given in the other three main windows: “Sat Pulse Data”, “N Status”, and

“Geospatial Data” (Table 3 to Table 5, respectively). The meaning of the status symbols is explained in Table 2. Measurements can only be started when one of the four main windows is displayed.


Table 1: LSA-2050 Window			
LSA-2050 Window			
Q ₃₁₀	0.13		
Q ₃₆₅	0.11		
Q ₄₅₀	0.81		
Q ₅₃₀	1.00		
C _{CHL}	86.6		
F _V /F _M	0.689		

Table 2: Status Indicators	
	Battery charge status. From left to right, full to low charge. Flashing, change batteries.
	LED Status. Left, visible ON. Right, UV ON. No Status LED, OFF.
	GPS status. Left, ON. Right, location determined.

In the N Status Window, the Q₃₆₅ and Q₅₃₀ values are converted into absorbance data (A_{FLAV} and A_{ANTH}, respectively). The NBI (nitrogen balance index) corresponds to the ratio of chlorophyll content to A_{FLAV} (see Section 6.4, page 36).

Table 3: N Status Window			
N Status Window			
NBI	61.2		
A _{FLAV}	0.82		
A _{ANTH}	0.07		
2024-03-20		15:28:41	

The F_v/F_M value of the window “LSA-2050” is also shown in the Sat Pulse Data window together with F_0 and F_M fluorescence levels ($F_v=F_M-F_0$), and the fluorescence transient induced by the saturation pulse.


Table 4: Sat Pulse Data	
Sat Pulse Data	
F_v/F_M	0.689
F_0	382
F_M	1230
	
2024-03-20	15:28:41

The window Geodata provides longitude and latitude (first line to the left, number followed by W or E; first line to the right, number followed by N or S, respectively), and height above sea level in meter. These position data is calculated from signals from GPS satellites which are picked up by the on-board GPS receiver. The number of available satellites is indicated in line 2 (#Sat). From position and world time (Coordinated Universal Time, UTC), the current sun azimuth (Sun Az.) and sun elevation (sun El.) is derived.

Leaf Azimut (Leaf Az.) and Leaf Slope (Leaf Sl.) are determined using gravity, magnetic field, acceleration and rotation data from the LSA-2050 instrument.

By combining the information on sun and leaf position, the angle of incidence is derived (A. o. I.). This number describes the angle at which sunlight strikes the leaf surface. The cosine of the angle of incidence is called surface incidence (Inc. %). The surface incidence is of relevance because the effective intensity of sun radiation at the leaf surface is porportional to this value.

Chapter 7.4 (page 51) summarizes all hardware involved in geodata acquisition. The same chapter explains the terminology used in geodata.

Table 5: Geodata			
Geodata			
12.345678 E		12.345678 N	
Height	400	#Sat	15
Leaf Az.	76	Sun Az.	135
Leaf Sl.	45	Sun El.	60
A. o. l.	60	Inc.	50
2024-03-20		17:41:43	

5.2.2 Main Menu

The Main Menu can be accessed from any of the three main windows by pressing the MENU button on the instrument.

Table 6: Main Menu	
Main Menu	
Settings	→
Calibration	→
Create New File	→
Memory	→
Device Info	→

5.2.2.1 Settings

The first item of the main menu leads to the menu Settings (Table 7). In any of the submenus of Settings, choose item by up and down keys. Then make selection by pressing the OK key.

Table 7: Settings	
Settings	
Active Tests	→
Chl. Settings	→
Mesophyll Type	→
Device Settings	→

In Settings, the "Active Tests" option allows you to disable tests. (Table 8). To save energy, or when battery power is low, switch off geodata. Geodata sensors consume a lot of energy.

Table 8: Active Tests	
Active Tests	
Chl. Concentration	on/off
F _v /F _M	on/off
Screening	on/off
Geodata	on/off

The item Chl. Settings allows changing the unit of chlorophyll concentration, and the calibration curve used to calculate chlorophyll concentrations (see Section 7.1, page 39). The selection of the unit for chlorophyll concentration does not affect the exported data: both nmol/cm² and µg/cm² are exported. Changing the calibration curve creates a new file with the next measurement.

Table 9: Chl. Settings	
Chl. Settings	
nmol/cm ²	X
µmol/cm ²	
C ₃ Type	X
C ₄ Type	

The mesophyll reference used to evaluate radiation screening can be selected in the Mesophyll Reference item (see 7.3, page 45). As in the case of calibration curves, changing the mesophyll reference creates a new file.

Table 10: Mesophyll Reference		
Mesophyll Reference		
Hylotelephium	Upper side	
Hylotelephium	Lower side	
Kalanchoe	Upper side	
Spinacia	Lower side	
Tulipa	Upper side	
Valeriana	Lower side	X

Device Settings configures the idle time after which the device is automatically switched off. You can also turn off the beep that accompanies keystrokes. Note that both the beginning and end of a measurement are signaled by a beep.

Table 11: Device Settings	
Device Settings	
Auto Off	15:00
Beeper	on/off
Load Defaults	→
Clear Memory	→

„Load Default” The default settings, which are activated by the command, are outlined in Section 5.1 (page 14).

The “Clear Memory” command erases all data without the option to recover them.




5.2.2.2 Calibration

Prior to measurements, three calibration procedures must be performed: Offset Values, Absorbance, and Fluorescence (Table 12). The fourth calibration (Orient. Sensor) is not needed for leaf analysis but for information on sun exposure of the leaf.

“Offset Values” determine the background signal of the device, with the sandwich light diffusing/black standard in place (see Fig. 1, page 7). “Absorbance” calibration measures the intensity of the FR and NIR LED in the presence of the Walz light diffusing standard. Fluorescence calibration determines the signal with the Walz fluorescence standard (see Section 7.3, page 45). All calibration values are documented in the exported data (Section 5.3.1, page 22).

Table 12: Calibration	
Calibration	
Offset Values	→
Absorbance	→
Fluorescence	→
Orient. Sensor	→

Sensors involved in determining the leaf position are calibrated in the window “Orient Sens. Calibration” (Table 13). Three orientation sensors are calibrated. The sensors and the meaning of the displayed numbers (Heading, Pitch, Role) are introduced in Section 7.4 (page 51).

Table 13: Orient. Sens. Calibration		
Orient. Sens. Calibration		
Gyro		Heading
Accel		Pitch
Mag		Roll
Various instruction texts.		

To calibrate the orientation sensors, follow the instructions displayed in the lower part of the window. Calibration of the gyroscope occurs first. Horizontal bars indicate the progress of calibration. Calibration switches automatically from gyroscope to accelerometer. In most cases, calibration of accelerometer and magnetometer occur simultaneously. When separate magnetometer calibration is required, you will be asked to describe the shape of the number 8, do so with the device tilted.

5.2.2.3 Create New File

The command Create New File will be executed with the next measurement.

5.2.2.4 Memory

The Memory item permits scrolling through stored measurements using the up and down keys. For each data set, the initial view is the LSA-2500 window. Pressing the Menu (➤) key sequentially shows the Windows “N Status”, “Sat Pulse”, and “Geo-data”. ESC (◀) returns to the previous view.

5.2.2.5 Device Info

The window “LSA-2050 Info” reports the charge status of the batteries. When connected to a computer, with batteries installed, the charge status indicates 100%. The device and software specific information is needed when reporting errors to the Walz staff.

Table 14:	LSA-2050 Info
LSA-2050 Info	
Battery: 80%	
S/N: LSA0101	
Firmware: 22/2311	
Build: 24-05-07 15:16:59	
Prog: 24-05-17 08:32:16	

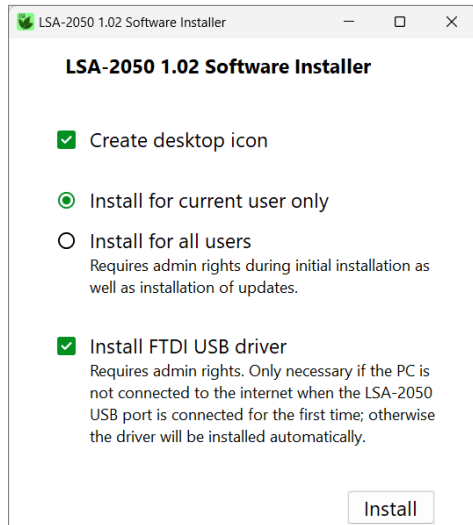
5.3 LSA-2050 Software

5.3.1 Installing the Software

Check the Walz website regularly for software updates: https://www.walz.com/products/chl_p700/leaf-state-analyzer/downloads.html. Always use the latest software!

Execute the LSA software installer (e.g., LSA-2050-v1.04-Installer.exe) located on the Walz USB flash drive by double clicking on the file. You can also copy the LSA installer to your computer and install it from there.

The software installation window opens (Fig. 8). Read the instructions carefully and select the appropriate option for your computer configuration. The FTDI USB driver is needed for communication with the LSA-2050 device. Click Install. A link to the LSA software is added to the program list in the Windows Start menu. Depending on choice, an LSA desktop icon is generated.

Fig. 8: Installation Window

5.3.2 Connecting the LSA-2050 to Computer

Remove cover of the battery compartment to gain access to the USB-C port of the LSA-2050 (Fig. 9).

Fig. 9: Access to USB-C Port

Connect to the computer using the Walz USB-C to USB-A cable (included in delivery) or a commercially available cable. Execute LSA-2050 software. Outdated firmware on the LSA-2050 device will automatically be updated. Then, the download window appears.

5.3.3 Adjusting Time

Click on the gear icon on the download window, to set your local time zone. Instructions are given in the legend of Fig. 10.

5.3.4 Downloading Data

The download window lists all the files that are stored on the LSA-2050 (Fig. 10, page 25). Left click once on the file to be downloaded. This action opens a file dialogue box. Choose file name and directory and click . A green checkmark indicates that the file has been successfully downloaded, that is, an Excel file has been created. A red cross indicates a download error.

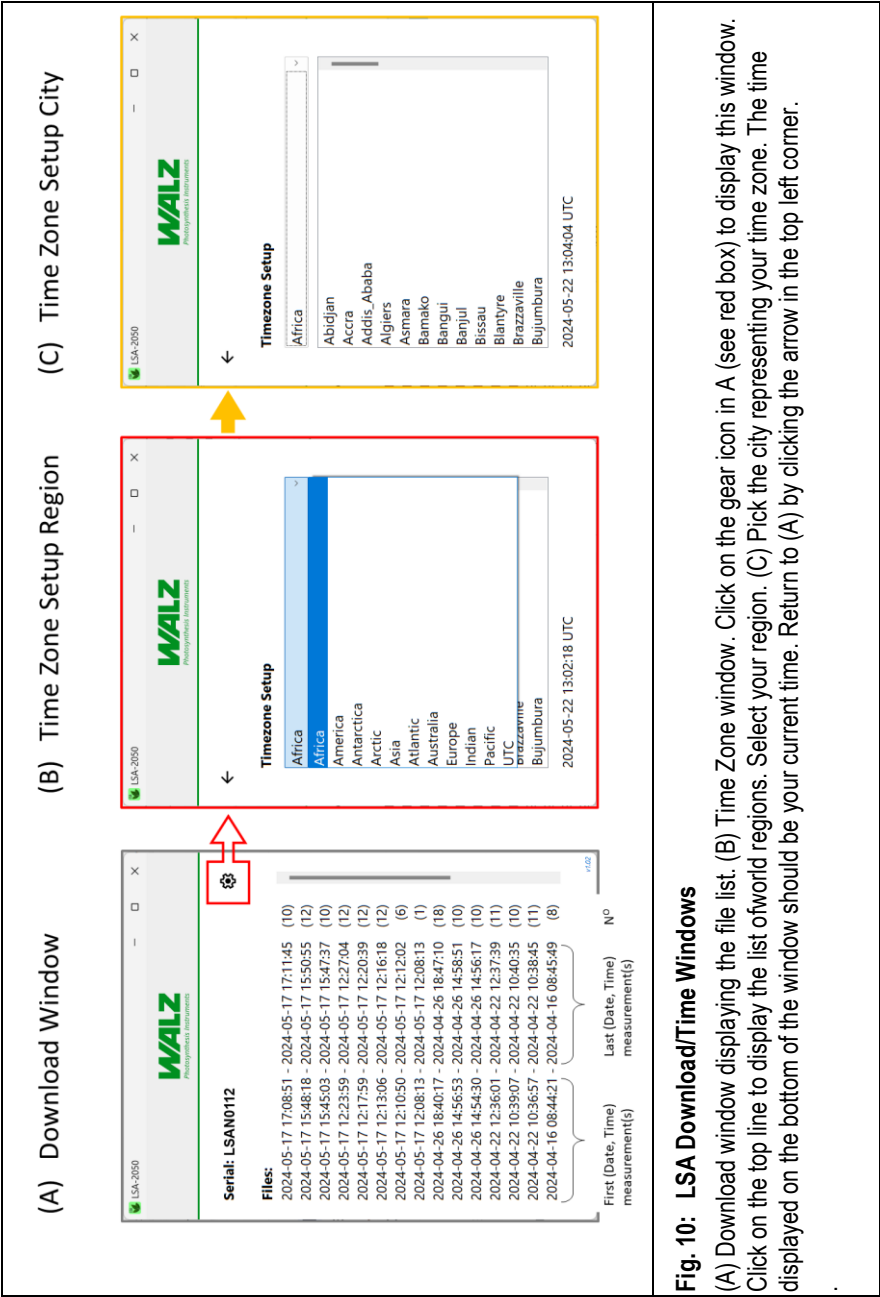


Fig. 10: LSA Download/Time Windows





(A) Download window displaying the file list. (B) Time Zone window. Click on the gear icon in A (see red box) to display this window. Click on the top line to display the list of world regions. Select your region. (C) Pick the city representing your time zone. The time displayed on the bottom of the window should be your current time. Return to (A) by clicking the arrow in the top left corner.

5.3.5 Excel File

The Excel file contains two sheets called “Measure” (Fig. 11, page 27) and “SAT Chart” (Fig. 12, Page 28). The initial view of the sheet “Measure” displays the basic sample properties and the incidence value (see Fig. 22, page 55). Table 15 (page 29) explains the abbreviations used.

The SAT Chart sheet contains the fluorescence transients from which the F_0 and F_M levels are derived (Fig. 12, page 28). Instructions on how to graph this data are given in the figure legend.

The sheet “Measure” contains all data used for the calculations of the sample properties, and of the incidence value. The buttons to display all data are highlighted by a green frame in Fig. 11.

The  button on the top right corner fully unfolds the Excel spreadsheet, the  above column R reveals all fluorescence data (Table 16, page 30 and Table 17, page 31), the  above column AD reveals all absorption data of chlorophyll determination, and the  above column AR reveals all geodata. All data are explained in Table 16, page 30 to Table 18, page 31.

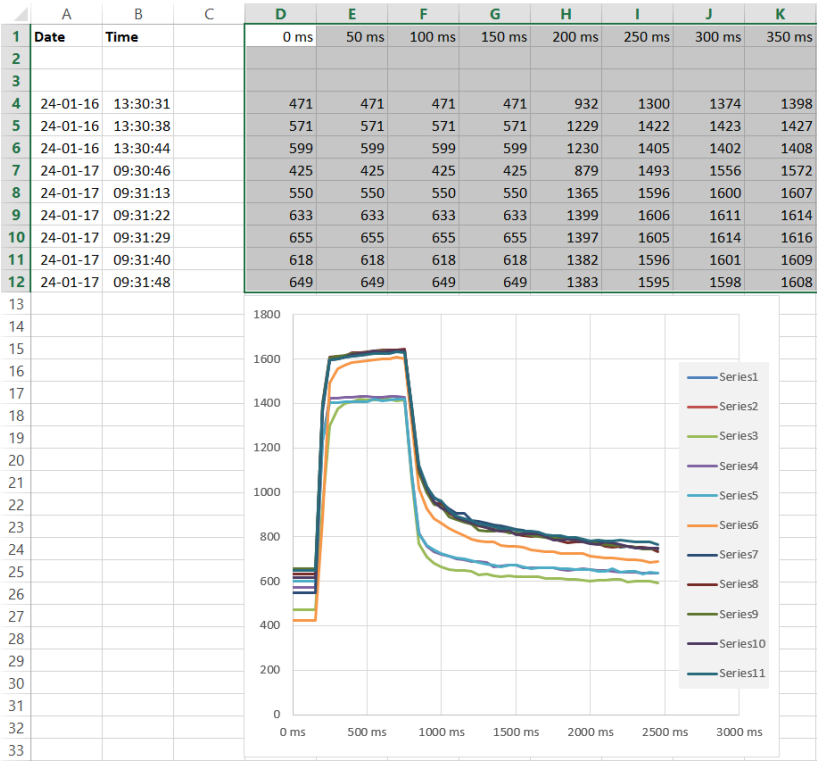


Fig. 12: SAT Chart

The “SAT Chart” contains the data of fluorescence transients used for F_V/F_M determinations. To graph the data, select time (row 1) and fluorecence data (row 4 to row n) from column D to column BA. The entire time row is conveniently selected by a left click in the first “time” cell (D1), followed by first simultaneously pressing Shift + Ctrl and then the right arrow key. Fluorecence data can be successively added by pressing the Shift key and then the down arrow key. All data can be selected by left click in cell D1, holding down the Shift + Ctrl, followed by pressing the right arrow key and the down arrow key. After data selection, go to window “Inser”, and pick “insert scatter (x, y) graph” in the section “Charts”.

Table 15: Results		
Column titles of the initial view of the Excel spreadsheet created by the LSA-2050 software (see Fig. 11, page 27)		
Column	Title	Comment
A	Date	Date in the format YY-MM-DD
B	Time	Time in the format hh:mm:ss
C	Type	Indicator of data source.
R	F_v/F_M	Maximum photochemical quantum yield of photosystem II.
T U V W	Q₃₁₀ Q₃₆₅ Q₄₅₀ Q₅₃₀	Apparent UV-B, UV-A, blue light, and green light screening, respectively. (Fluorescence excited at 310 nm, 365 nm, 450 nm, and 530 nm, respectively, relative to fluorescence excited at 630 nm.)
X	A_{FLAV}	The A _{FLAV} (absorbance by flavonoids) is derived from Q ₃₆₅ and can be considered as proportional to the concentration of flavonoids involved in UV-A screening.
Y	A_{ANTH}	The A _{ANTH} (absorbance by anthocyanins) is derived from Q ₅₃₀ and can be considered as proportional to the concentration of anthocyanins involved in green light screening.
AE	nmol/cm²	Concentration of Chl <i>a</i> + Chl <i>b</i>
AF	µg/cm²	Concentration of Chl <i>a</i> + Chl <i>b</i>
AG	NBI	Nitrogen balance index.
AR	Incidence	Cosine of the angle at which solar radiation hits the leaf.

Table 16: Calibration Data		
Row	Title	Comment
2 Offset	I_{310} I_{Fo} I_{365} I_{Fm} I_{450} I_{530} I_{700} I_{615} I_{770}	Background signal (Offset) for all LEDs and methods (system property determined by calibration).
3 Reference	I_{310} F_{310} I_{365} F_{365} I_{450} F_{450} I_{530} F_{530} I_{615} F_{615}	I_{λ} : Original signal at wavelength λ measured with the fluorescence standard. F_{λ} : I_{λ} corrected by the background signal.
3 Reference	Q_{310} Q_{365} Q_{450} Q_{530}	Mesophyll reference factors (see 7.3.1, page 47).
2, 3, and 4	nmol/cm^2 $\mu\text{g/cm}^2$	Coefficients for calculating the molar and weight concentrations of Chl <i>a</i> + Chl <i>b</i> (see Section 7.1.4, page 42). In column AD, "C3" and "C4" indicate which calibration curve was used to calculate concentrations; "Coeff Q" is the coefficient of the quadratic term, "Coeff L" is the coefficient of the linear term of the calibration curve, and "Coeff C" is a constant (See Section 7.1, page 39).

Table 17: Source Data 01		
Data from row 4 onwards		
Title		Comment
I_{F0} I_{Fm}	F₀ F_m	I _{F0} , raw fluorescence signal of the dark leaf. I _{Fm} , raw fluorescence signal of the leaf exposed to a strong (saturating) light pulse. F ₀ and F _m are the corresponding signals after offset correction. F _v /F _M in Table 15 (page 29) equals 1-F ₀ /F _m .
I₃₁₀ I₃₆₅ I₄₅₀ I₅₃₀ I₆₁₅	F₃₁₀ F₃₆₅ F₄₅₀ F₅₃₀ F₆₁₅	I _λ : Signal at wavelength λ measured with the leaf. F _λ : I _λ corrected by the background signal.
I₇₀₀ I₇₇₀		Signal induced by radiation transmitted by the leaf at 700 nm and 770 nm, respectively.
Transmittance		I ₇₀₀ /I ₇₇₀ of the sample normalized to I ₇₀₀ /I ₇₇₀ of calibration. All data was offset-corrected prior to calculation.
Absorbance		-log ₁₀ Transmittance.

Table 18: Source Data 02	
Data from row 4 onwards	
Title	Comment
Satellite #	Number of GPS satellites.
DoP	Dilution of precision. The number indicates the precision with which the GPS determines a position (<1, high precision. >20, very poor confidence).
Latitude Longitude	Coordinate parameters for a point on the Earth surface given in decimal degrees.
Height	Height above sea level in meters.
Leaf Az.	Leaf azimuth. See Fig. 21, page 54.
Leaf Sl.	Leaf slope. See Fig. 20, page 54.
Sun Az.	Sun azimuth. See Fig. 19, page 53.
Sun Elev.	Sun elevation. See Fig. 19, page 53.
Aol	Angle of incidence. See Fig. 22, page 55.
Incidence	Cosine of angle of incidence. See Fig. 22, page 55.

6 Comments on Results

This section shortly treats meaning and interpretation of the parameters obtained with the LSA-2050 LEAF-STATE-ANALYZER. Cited literature is listed right after the text.

6.1 F_V/F_M

Kitajima and Butler (1975) have measured at low temperature (77 K), the effect of a fluorescence-quenching chemical on the fluorescence ratio of F_V/F_M . They have mathematically derived that the F_V/F_M is a measure for the maximum yield for primary photochemistry of photosystem II. With the same technique, Björkman and Demmig (1987) have determined this value to be around 0.83 in many species with C_3 photochemistry.

This finding has been confirmed by PAM fluorescence, and it has additionally been shown that the F_V/F_M of plants having C_4 photochemistry can be as low as 0.76 (Pfündel 1998, Pfündel et al. 2013). Guidi et al. (2019) have provided a compilation of F_V/F_M values in various species.

When F_V/F_M values are significantly lower than the maximum F_V/F_M , photoinhibition of photosystem II is likely (Maxwell and Johnson 2000). This means that a part of the photosystem II in the sample has damaged reaction centers.

For correct F_V/F_M measurements, it is important that leaves are fully dark-acclimated. Depending on species and growth conditions, dark times can range between 10 minutes and many hours. The dark time for F_V/F_M measurements must be determined experimentally.

Björkman O, Demmig B (1987) Photon yield of O₂ evolution and chlorophyll fluorescence characteristics at 77 K among vascular plants of diverse origins. *Planta* 170, 489–504 (1987). <https://doi.org/10.1007/BF00402983>

Guidi L, Lo Piccolo E, Landi M (2019) Chlorophyll fluorescence, photoinhibition and abiotic stress: does it make any difference the fact to be a C3 or C4 Species? *Front Plant Sci* 10:174. <https://www.frontiersin.org/journals/plant-science/articles/10.3389/fpls.2019.00174/full>

Kitajima M, Butler WL (1975) Quenching of chlorophyll fluorescence and primary photochemistry in chloroplasts by dibromothymoquinone. *Biochim Biophys Acta – Bioenergetics* 376: 105–115. [https://doi.org/10.1016/0005-2728\(75\)90209-1](https://doi.org/10.1016/0005-2728(75)90209-1)

Maxwell K, Johnson GN (2000) Chlorophyll fluorescence—a practical guide, *Journal of Experimental Botany* 51: 659–668. <https://doi.org/10.1093/jexbot/51.345.659>

Pfündel E (1998) Estimating the contribution of Photosystem I to total leaf chlorophyll fluorescence. *Photosynthesis Research* 56, 185–195. <https://doi.org/10.1023/A:1006032804606>

Pfündel EE, Klughammer C, Meister A, Cerovic ZG (2013) Deriving fluorometer-specific values of relative PSI fluorescence intensity from quenching of F_0 fluorescence in leaves of *Arabidopsis thaliana* and *Zea mays*. *Photosynth Res* 114, 189–206. <https://doi.org/10.1007/s11120-012-9788-8>

6.2 Screening

The fundamental articles of radiation-screening substances in the leaf are listed in Section 7.3, page 45. As has been introduced by Bilger et al. (1997), the LSA-2050 outputs fluorescence quotients (Q_λ) as a measure for the extent of screening. The quotient ranges from 0 to 1, where 1 signifies full transmittance (no screening).

As a rule of thumb, leaves are sufficiently protected against UV-B radiation when the Q_{310} is 0.1 or smaller. Similar efficient screening has been reported for green light screening (Q_{530}) by anthocyanins (Nichelmann and Bilger 2017).

The fluorescence quotients (Q_λ) can be viewed as transmittance values. Transmittance can be converted to absorbance, which

ideally is proportional to the concentration of screening compounds (Goulas et al. 2004).

The LSA-2050 converts the quotients Q_{365} and Q_{530} into absorbance data which are supposed to be proportional to the concentration of flavonoids and anthocyanins, respectively. These absorbance values reflect the total concentration in the leaf only if the screening pigments are almost exclusively located in the epidermis which is probed. This requirement seems to be met, e.g., in grapevine but not in barley (Kolb and Pfündel 2005).

Kolb CA, Pfündel EE (2005) Origins of non-linear and dissimilar relationships between epidermal UV absorbance and UV absorbance of extracted phenolics in leaves of grapevine and barley. *Plant, Cell & Environment* 28: 580-590.

<https://doi.org/10.1111/j.1365-3040.2005.01302.x>

Nichelmann L, Bilger W (2017) Quantification of light screening by anthocyanins in leaves of *Berberis thunbergii*. *Planta* 246: 1069–1082 (2017). <https://doi.org/10.1007>

6.3 Chlorophyll Concentration

Leaf chlorophyll concentrations respond to nutrient availability and various stress factors including pollution or herbivory (Agathokleous et al. 2020). Relationships between leaf chlorophyll and the leaf nitrogen status have been shown (Evans 1989, Lu et al. 2020, Xiong et al. 2015). That nitrogen fertilization can elevate leaf contents of chlorophyll and nitrogen has been demonstrated (Prsa et al. 2007, Muhammad et al. 2022).

In summary, the leaf chlorophyll concentration can reflect stress effects and insufficient nitrogen supply. Because the leaf chlorophyll concentration varies between species, and the relationships between chlorophyll and nitrogen are variable, universal chlorophyll values for healthy plant cannot be given. However, by analyzing comparable plants after different treatments, the

measurement of chlorophyll concentration becomes a highly meaningful stress detector.

Agathokleous E, Feng ZZ, Peñuelas J (2020) Chlorophyll hormesis: Are chlorophylls major components of stress biology in higher plants? *Sci Total Environ* 726, 138637. <https://doi.org/10.1016/j.scitotenv.2020.138637>

Evans JR (1989) Photosynthesis and nitrogen relationships in leaves of C3 plants. *Oecologia* 78: 9-19. <https://doi.org/10.1007/bf00377192>

Lu X, Ju W, Li J, Croft H, Chen JM, Luo Y, Yu H, Hu H (2020). Maximum carboxylation rate estimation with chlorophyll content as a proxy of Rubisco content. *J Geophys Res Biogeosci* 125: e2020JG005748. <https://doi.org/10.1029/2020JG005748>

Muhammad I, Yang L, Ahmad S; Farooq S, Al-Ghamdi AA, Khan A, Zeeshan M, Elshikh MS, Abbasi AM, Zhou X-B (2022) Nitrogen fertilizer modulates plant growth, chlorophyll pigments and enzymatic activities under different irrigation regimes. *Agronomy* 12: 845. <https://doi.org/10.3390/agronomy12040845>

Prsa I, Stampar F, Vodnik D, Veberic R (2007) Influence of nitrogen on leaf chlorophyll content and photosynthesis of 'Golden Delicious' apple. *Acta Agric Scand B Soil Plant Sci* 57: 283-289. <https://doi.org/10.1080/09064710600982878>

Xiong D, Chen J, Yu T, Gao W, Ling X, Li Y, Peng S, Huang J (2015). SPAD-based leaf nitrogen estimation is impacted by environmental factors and crop leaf characteristics. *Sci Rep* 5: 13389. <https://doi.org/10.1038/srep13389>

6.4 Nitrogen Balance Index (NBI),

The NBI has been defined as the concentration ratio of chlorophyll to UV-absorbing phenolics, mainly flavonoids (see papers below). The NBI was formulated on the basis that the chlorophyll content increased with nitrogen supply and the phenolic content tended to decrease in parallel. Because of these opposing relationships, the NBI potentially responds more sensitive and robust to the plant nitrogen status than the individual concentrations of chlorophyll and phenolics.

The NBI is expected to increase with nitrogen supply. The relationship between NBI and nitrogen status, however, may vary between species and even cultivars of the same species. Therefore, universal NBI values indicating optimum nitrogen supply are not available but must be experimentally established. However, once the NBI for optimal nitrogen supply has been established for a plant culture, the NBI can be a valuable tool for matching fertilization to crop requirements.

Cartelat A, Cerovic ZG, Goulas Y, Meyer S, Lelarge C, Prioul J-L, Barbottin A, Jeufroy M-H, Gate P, Agati G, Moya I (2005) Optically assessed contents of leaf polyphenolics and chlorophyll as indicators of nitrogen deficiency in wheat (*Triticum aestivum* L.). *Field Crops Res* 91: 35–49. <https://doi.org/10.1016/j.fcr.2004.05.002>

Cerovic ZG, Ben Ghazlen N, Milhade C, Obert M, Debuissou S, Le Moigne M (2015) Nondestructive Diagnostic test for nitrogen nutrition of grapevine (*Vitis vinifera* L.) based on Dualex leaf-clip measurements in the field. *J Agric Food Chem* 63 (14): 3669–3680. <https://doi.org/10.1021/acs.jafc.5b00304>

7 Documentation

7.1 Chlorophyll Calibration

7.1.1 Plant Species

Four plant species were investigated (Table 19). The species were chosen to represent monocot (*T. aestivum*, *Z. mays*) and dicot leaf anatomy (*H. annuus*, *N. tabacum*). The species also represent different types of photosynthesis with different Chl *a*/Chl *b* concentration ratios (Table 19).

Table 19: Plant Species used for Chl Calibration

	Species	Group	Type of Photosynthesis	Chl <i>a</i> /Chl <i>b</i>
A	Common sunflower (<i>Helianthus annuus</i>) Asteraceae	Dicot	C ₃	Normal
B	Common wheat (<i>Triticum aestivum</i>) Poaceae	Monocot	C ₃	Normal
C	Tobacco (<i>Nicotiana tabacum</i>) Solanaceae	Dicot	C ₃	Normal
D	Corn (<i>Zea mays</i>) Poaceae	Monocot	C ₄ NADP-ME	Elevated

7.1.2 Growth Conditions

In case of the species *H. annuus*, *T. aestivum*, and *Z. mays*, (A, B, and D, in Table 19) varying chlorophyll contents were achieved by five different growth conditions (Table 20). In another experiment, leaves of *N. tabacum* cv. Samsun NN were probed. These plants had senescing leaves with very low chlorophyll contents.

Table 20: Growth Conditions

<i>H. annuus</i> , <i>T. aestivum</i> , and <i>Z. mays</i>			
Treatment	Fertilization*	Additional Lighting**	Location
1	None	Yes	Greenhouse
2	1 g / pot	Yes	Greenhouse
3	3 g / pot	Yes	Greenhouse
4	None	No	Greenhouse
5	3 g / pot	No	Greenhouse→Field***

*40 days after sowing (June 13, 2022): Compound fertilizer (“Blaukorn”) added, consisting of 12% N, 8% P₂O₅, 16% K₂O, 3% MgO, 23% SO₃, 0.02% B, 0.06% Fe, 0.01% Zn. Pots: 17.5 cm x 17.5 cm x 17 cm containing 4.5 liter soil (compost : sand : clay = 2:1:1).

**400 W high-pressure sodium vapor lamps mounted 100 cm above the planting table, providing about 250 μmol m⁻² s⁻¹ from 06:00 until 18:00, additionally to natural PPFD.

***42 days after sowing (June 15, 2022): Moving to the field.

7.1.3 Chl. Measurements

In the first experiment (species A, B, and D, in Table 19), measurements took place 55 days after sowing (June 28, 2022) in dim light. First, the spots to be measured were marked. Then, attached leaves were sequentially probed with three prototypes

of the LSA-2050 using the wavebands introduced by Cerovic et al. (2012).

The marked spots were then punched out with a 1 cm diameter cork borer and the leaf disks were frozen in liquid nitrogen. The frozen disks were shipped on dry ice to the laboratory where they were stored at -80°C until pigment analysis.

Photosynthetic pigments were extracted with 100% acetone as described in Bethmann et al. (2019):

Individual pigments were separated and quantified by high-performance liquid chromatography (HPLC, Fig. 13) according to Färber et al. (1997).

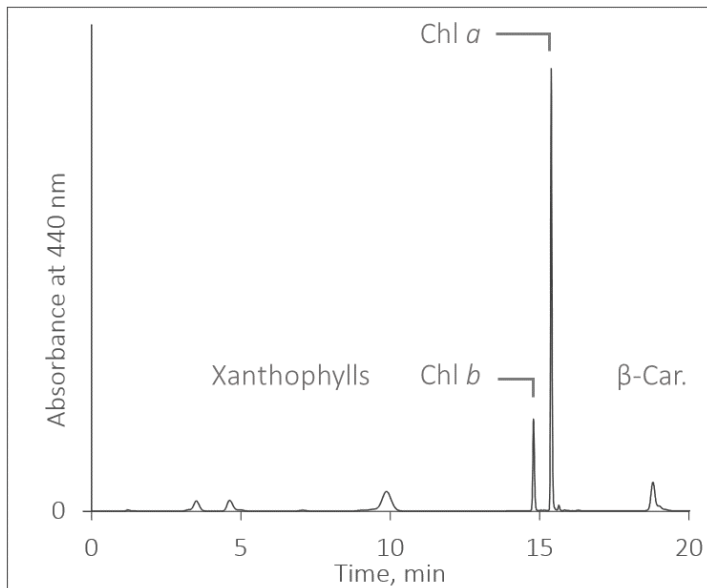


Fig. 13: Chromatogram for Pigment Determination

In *N. tabacum*, pigment extraction was as the forgoing experiment, but chlorophyll quantification was spectrometrically according to Lichtenthaler and Buschmann (2001).

7.1.4 Correlations

The data of the three species with C_3 photosynthesis could be described by a single function. Curvilinear relationships with high coefficients of determination were observed for the relationships between total chlorophyll content (Chl *a* + Chl *b*) and absorbance measured by the LSA-2050 device (Table 21).

The relationship of the C_4 plant differed from that of the C_3 species: the same absorbance value yielded a lower Chl *a* + Chl *b* concentration compared to the relationship for C_3 plants. This may originate in relatively low abundance of Chl *b* in the C_4 plant and the fact that the LSA-2050 measures mainly Chl *a* light absorption (compare Fig. 14). Also, the optical properties of the C_4 Kranz anatomy could have affected the correlation for *Z. mays*.

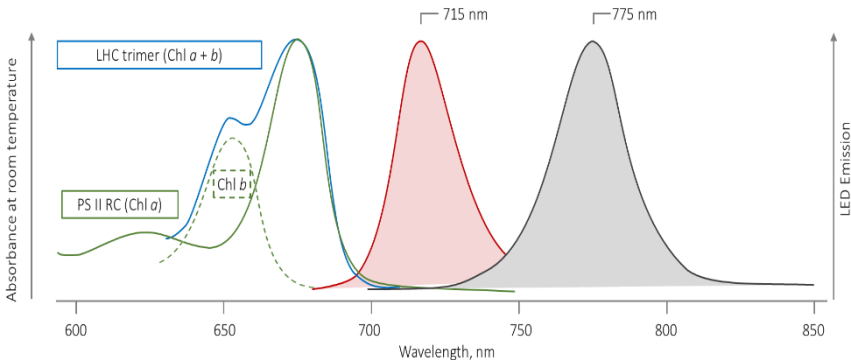


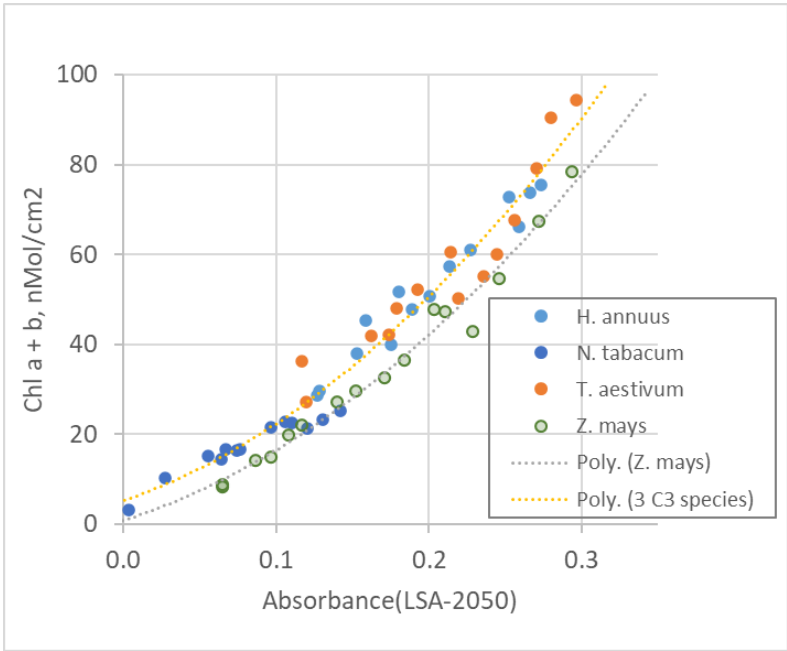
Fig. 14: Chlorophyll determination wavelengths

The LSA-2050 mainly measured Chl *a* light absorption because the Chl *b* absorbance spectrum does not overlap meaningfully with the emission spectrum of the sample LED. Red line and pink-filled: emission spectrum of the sample LED. Dark-grey line and grey-filled: emission spectrum of the reference LED. Green line: absorbance spectrum of the photosystem II reaction center containing only Chl *a*. Blue line: absorbance spectrum of the trimer of the light harvesting complex of photosystem II containing Chl *a* and Chl *b*. Green dashed line: approximate absorption spectrum of Chl *b*. Absorbance spectra of photosynthetic complexes redrawn after Mendes-Pinto et. al (2013)

Table 21: Chl. Correlations

Species	Chl a + b, nMol/cm ²	Chl a + b, µg/cm ²
<i>H. annuus</i> , <i>N. tabacum</i> , <i>T. aestivum</i>	$y = 565x^2 + 115x + 5$ $R^2 = 0,964$	$y = 505x^2 + 102x + 5$
<i>Z. mays</i>	$y = 501x^2 + 106x + 1$ $R^2 = 0.981$	$y = 449x^2 + 95x + 1$

x=absorbance as measured by an LSA-2050.



Bethmann S, Melzer M, Schwarz N, Jahns P (2019) The zeaxanthin epoxidase is degraded along with the D1 protein during photoinhibition of photosystem II. *Plant Direct* 3: 1–13. <https://doi.org/10.1002%2Fpld3.185>

Cerovic ZG, Masdoumier G, Ben Ghazlen N, Latouche G (2012) A new optical leaf-clip meter for simultaneous non-destructive assessment of leaf chlorophyll and epidermal flavonoids. *Physiol Plant* 146: 251–260. <https://doi.org/10.1111%2Fj.1399-3054.2012.01639.x>

Färber A, Young AJ, Ruban AV, Horton P, Jahns P (1997) Dynamics of xanthophyll-cycle activity in different antenna subcomplexes in the photosynthetic membranes of higher plants (The relationship between zeaxanthin conversion and nonphotochemical fluorescence quenching). *Plant Physiol* 115: 1609–1618. <https://doi.org/10.1104/pp.115.4.1609>

Lichtenthaler HK, Buschmann C (2001) Chlorophylls and carotenoids - Measurement and characterisation by UV-VIS. *Current Protocols in Food Analytical Chemistry* (CPFA), John Wiley, New York, 2001, Supplement 1 pp. F4.3.1eF4.3.8.

Mendes-Pinto MM, Galzerano D, Telfer A, Pascal AA, Robert B, Illoiaia C (2013) Mechanisms underlying carotenoid absorption in oxygenic photosynthetic proteins. *Biol Chem* 288: 18758 –18765. <https://doi.org/10.1074%2Fjbc.M112.423681>

Mendes-Pinto MM, Galzerano D, Telfer A, Pascal AA, Robert B, Illoiaia C (2013) Mechanisms underlying carotenoid absorption in oxygenic photosynthetic proteins. *Biol Chem* 288: 18758 –18765. <https://doi.org/10.1074%2Fjbc.M112.423681>

7.2 Chlorophyll and Small Leaves

To measure chlorophyll concentration on small leaves, the emitter aperture can be reduced to a spot of 6 mm diameter without affecting the chlorophyll measurements.

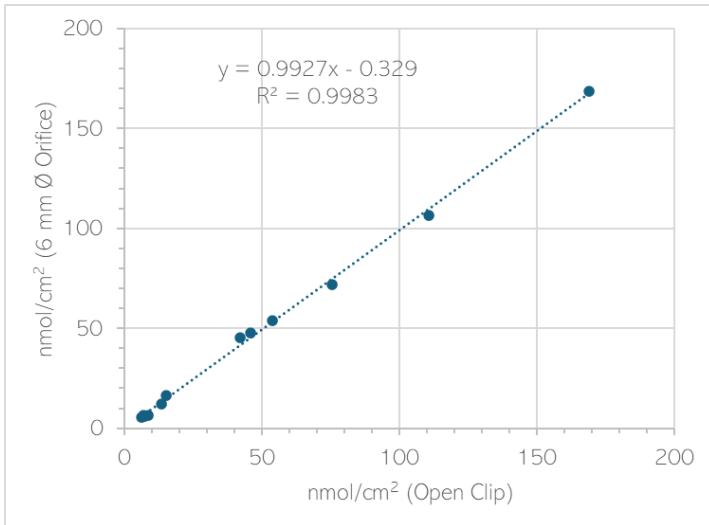


Fig. 15: Chlorophyll Determination with Small Leaves

Various samples were probed with the default configuration of the LSA-2050, and then again with the emitter opening reduced to a spot of 6 mm diameter. The graph shows the latter data plotted against the corresponding former data. The straight line, its equation and the R^2 result from linear regression.

7.3 Screening

Screening of photosynthetic pigment-protein complexes against UV-B and UV-A radiation by phenolics was non-invasively measured according to:

Bilger W, Veit M, Schreiber L, Schreiber U (1997) Measurement of leaf epidermal transmission of UV radiation by chlorophyll fluorescence. *Physiol Plant* 101: 754–763. <https://doi.org/10.1111/j.1399-3054.1997.tb01060.x>

Cerovic and coworkers have extended the Bilger method into the visible range by using green light to assess light screening by anthocyanins.

Cerovic ZG, Moise N, Agati G, Latouche G, Ben Ghazlen N, Meyer S (2008) New portable optical sensors for the assessment of winegrape phenolic maturity based on berry fluorescence. *Journal of Food Composition and Analysis* 21: 650-654.

<https://doi.org/10.1016/j.jfca.2008.03.012>

Nichelmann and coworkers have taken a further step by introducing blue light to probe screening by carotenoids that are not or only partially active in photosynthetic light harvesting.

Nichelmann L, Schulze M, Herppich WB, Bilger W (2016) A simple indicator for non-destructive estimation of the violaxanthin cycle pigment content in leaves. *Photosynthesis Research* 128: 183-193. <https://doi.org/10.1007/s11120-016-0218-1>

The LSA-2050 integrates these three approaches by measuring screening as apparent transmittance at UV-B, UV-A, blue, and green wavelengths with red as reference beam. For UV-A and green radiation, transmittance is converted into absorbance, to provide a relative number for the concentration of flavonoids and anthocyanins, respectively, which are active in light screening. Converting transmittance into absorbance for obtain a proxy for phenolics concentration has been introduced by Goulas et al. (2004):

Goulas Y, Cerovic ZG, Cartelat A, Moya I (2004) Dualex: a new instrument for field measurements of epidermal ultraviolet absorbance by chlorophyll fluorescence.

Appl Opt 43, 4488-4496. <https://doi.org/10.1364/ao.43.004488>

The LED combinations used together with the main classes of pigments absorbing at probing wavelengths are compiled in Table 22. The LED emission spectra are shown in Fig. 16.

LED Pair	Main class of screening pigments
310 nm vs. 630 nm	Hydroxycinnamic acids
365 nm vs. 630 nm	Flavonoids
450 nm vs. 630 nm	Carotenoids
530 nm vs. 630 nm	Anthocyanins

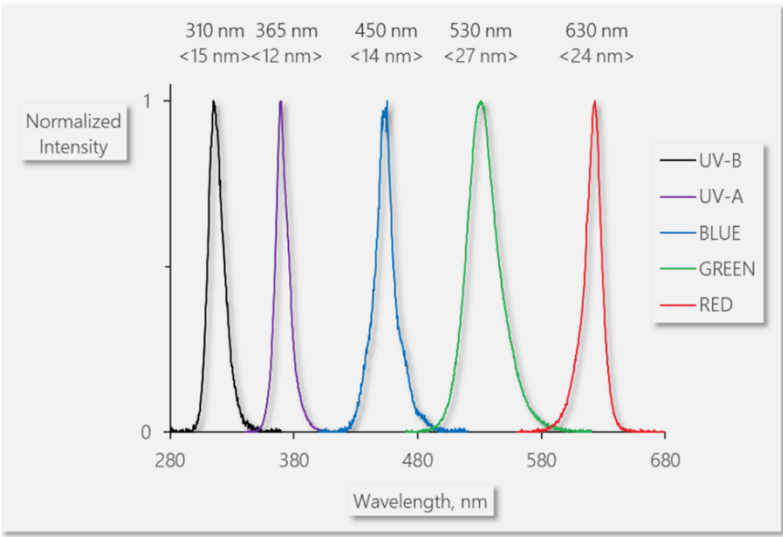


Fig. 16: Emission spectra of LEDs involved in screening measurements

Emission spectra normalized to the same maximum are shown. For each spectrum, peak wavelength, and full width at half maximum (<FWHM>) is indicated.

7.3.1 Mesophyll Reference Factor $RF(\lambda)$

To estimate the radiation screening of the leaf mesophyll, measurements of intact leaves must be related to measurements of unscreened mesophyll tissue. For this, five plant species with removable epidermis were selected (Table 23, page 49). For *H.*

telephium, both the upper and the lower leaf sides were probed. For the other species, either the upper or the lower leaf side was probed.

For all cases, mesophyll ratios, $MR(\lambda)_{MES}$, were established as defined by Eq. 1.

$$MR(\lambda)_{MES} = (I(\lambda)/I(630))_{MES} \quad \text{Eq. 1}$$

where $I(\lambda)$ is the fluorescence intensity excited at wavelength λ , and $I(630)$ is the fluorescence intensity excited at 630 nm. The λ represents one of the four wavebands: UV-B, UV-A, blue, or green (Fig. 16, page 47).

The same ratios were also established for the fluorescence standard of the LSA-2050. Dividing the mesophyll ratio by the corresponding standard ratio yields the mesophyll reference factor, $RF(\lambda)$:

$$RF(\lambda) = \frac{(I(\lambda)/I(630))_{MES}}{(I(\lambda)/I(630))_{STD}} \quad \text{Eq. 2}$$

The Reference Factor $RF(\lambda)$ is device independent. This is because different intensities of the LSA-2050 LEDs affect the intensity ratios measured with mesophyll and with the standard in the same way.



Fig. 17: Leaf of *H. telephium* with Epidermis Partially Removed

MES, free mesophyll. EP, stripped epidermis.

Table 23: Mesophyll Reference Factors

Mean values and 95% confidence intervals (95% CI) are given. $RF(\lambda)$ is defined by Eq. 2. In the LSA-2050, the $RF(\lambda)$ is denoted as “Mesophyll Reference”. The $RF(\lambda)$ can be selected via Main Menu→Settings→Mesophyll Type. In the Excel export file, the $RF(\lambda)$ values are written in cells T3 to W3.

Species Family	Leaf side	n		$RF(310)$	$RF(365)$	$RF(450)$	$RF(530)$
<i>Hylotelephium telephium</i> Crassulaceae	Upper	51	Mean	0.573	1.105	1.055	0.809
			95% CI	0.026	0.024	0.015	0.006
<i>Hylotelephium telephium</i> Crassulaceae	Lower	30	Mean	0.863	1.285	1.160	0.807
			95% CI	0.048	0.040	0.017	0.008
<i>Kalanchoe daigremontiana</i> Crassulaceae	Upper	10	Mean	0.689	1.208	0.937	0.844
			95% CI	0.061	0.041	0.030	0.047
<i>Spinacia oleracea</i> Amaranthaceae	Lower	10	Mean	0.738	1.146	1.146	0.734
			95% CI	0.072	0.040	0.038	0.045
<i>Tulipa spec.</i> Liliaceae	Upper	40	Mean	0.690	0.852	1.023	0.807
			95% CI	0.039	0.049	0.012	0.007
<i>Valeriana locusta</i> Caprifoliaceae	Lower	16	Mean	0.875	1.315	1.290	0.688
			95% CI	0.055	0.053	0.064	0.017

7.3.2 Calibration and Measurement

The calibration with the Walz fluorescence standard determines all four fluorescence ratios $(I(\lambda)/I(630))_{STD_DEVICE}$. These ratios are device-dependent because they are affected by the emission intensities of the LEDs.

Multiplying a ratio $(I(\lambda)/I(630))_{STD_DEVICE}$ with the corresponding $RF(\lambda)$ results in the “mesophyll reference” value $MR(\lambda)_{DEVICE}$. The $MR(\lambda)_{DEVICE}$ corresponds to the fluorescence ratio of the naked mesophyll adapted to the current LED intensities of the LSA-2050 instrument used.

$$MR(\lambda)_{DEVICE} = \left(\frac{I(\lambda)}{I(630)} \right)_{STD_DEVICE} \cdot RF(\lambda) \quad \text{Eq. 3}$$

The apparent screening is then the fluorescence ratio obtained with the sample, $(I(\lambda)/I(630))_{SAMPLE}$ divided by the corresponding mesophyll reference value:

$$Q(\lambda)_{SAMPLE} = \frac{(I(\lambda)/I(630))_{SAMPLE}}{MR(\lambda)_{DEVICE}} \quad \text{Eq. 4}$$

Calculations of $Q(\lambda)$ uses the $RF(\lambda)$ selected (compare Table 23).

Among all mesophyll references, *Valeriana locusta* exhibited the highest $RF(310)$ and $RF(365)$ (Table 23, page 49). The lower corresponding $RF(\lambda)$ of the other samples may arise from the presence of UV screening within the mesophyll. Therefore, the LSA-2050 uses the $RF(\lambda)$ values from *Valeriana locusta* as a default.

7.4 Geodata

Fig. 18 gives an overview of the components involved in providing geospatial information.

The GPS receiver obtains signals from satellites of the Global Positioning System. From satellite positions, latitude and longitude of the current position on Earth is calculated. The internal clock is set to UTC (Coordinated Universal Time) by an external timer like the Windows operating system. From the current position and UTC, the sun's position relative to ground is determined (compare Fig. 19, 53).

The magnetometer determines north based on the earth's magnetic field. The accelerometer detects gravity and changes in velocity in XYZ directions, and the gyroscope measured rotations in these three directions. By integrating the data of these three sensors, the leaf position (slope and azimuth) is calculated. Leaf slope and azimuth are illustrated in Fig. 20 (page 54) and Fig. 21 (page 54), respectively.

From the positions of sun and leaf, the angle at which sun radiation hits the leaf is derived (angle of incidence, Fig. 22, page 55). The physiologically relevant number is the cosine of the angle of incidence, because it indicates the relative effective intensity of sun radiation at the leaf surface.

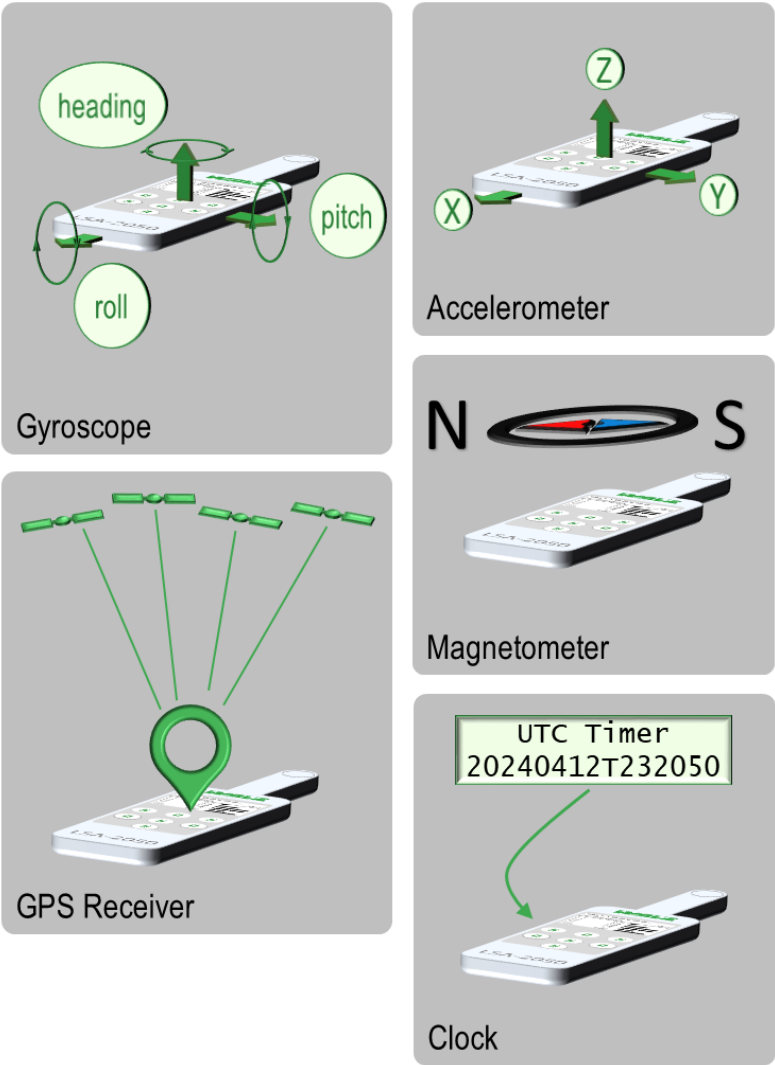


Fig. 18: Geodata Hardware

Devices and ways of geodata input. The gyroscope and the accelerometer measure rotation and velocity changes, respectively, in X, Y, and Z direction. The magnetometer records the earth magnetic field. The GPS receiver determines latitude and longitude of the current position. The UTC timer provides the world standard time.

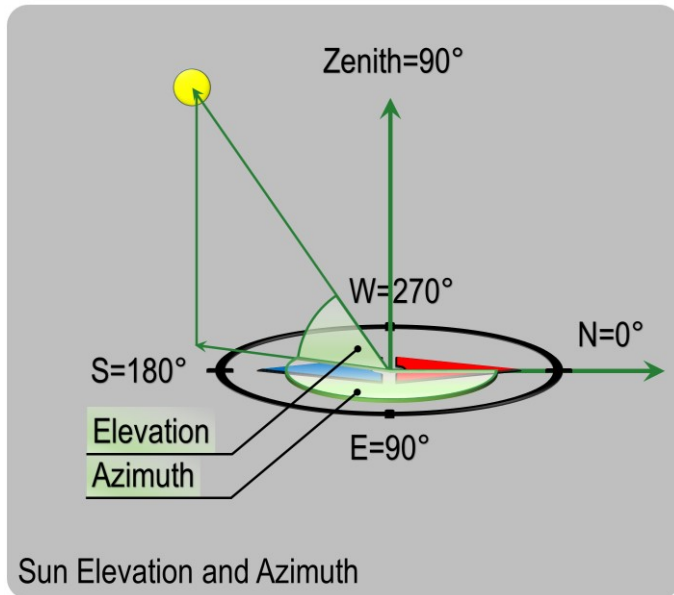


Fig. 19: Sun Position

The position of the sun relative to the Earth surface can be described by azimuth and elevation. Azimuth is the compass direction of the sun relative to the northern direction. With north being the zero point, the azimuth rises from 0° to 360° clockwise. Elevation is the sun's position above the horizon ranging from 0° (horizon) to 90° (zenith).

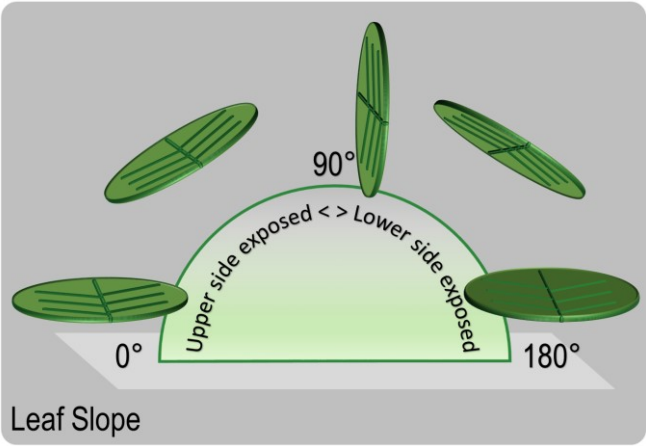


Fig. 20: Leaf Slope

The leaf slope is the leaf angle relative to the ground. 0° corresponds to the horizontal position of the leaf with top side up. 90° describes the vertical position and 180° is the horizontal position with the lower side up.

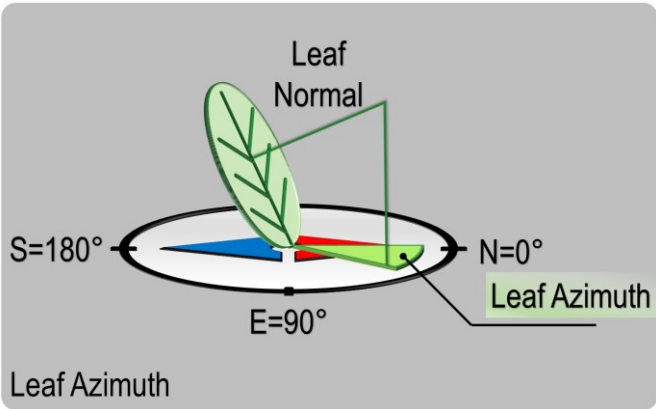


Fig. 21: Leaf Azimuth

The leaf azimuth is the compass direction of leaf normal relative to the northern direction. The range of the leaf azimuth is identical to that of the sun azimuth.

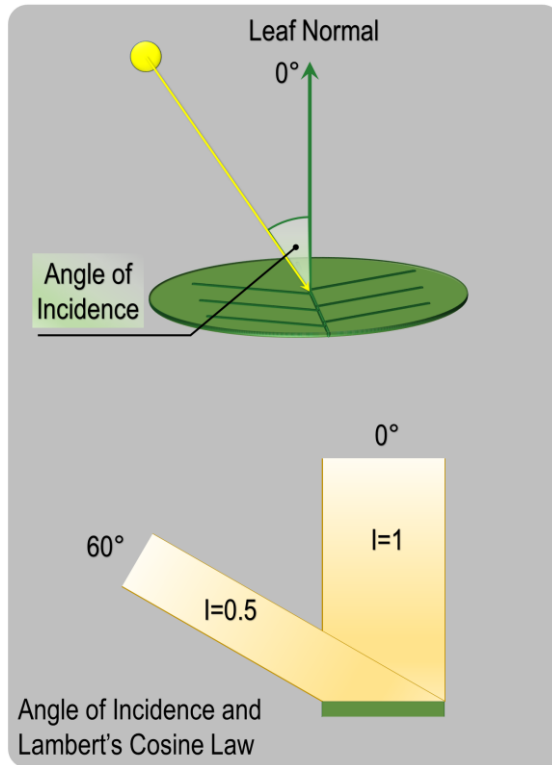


Fig. 22: Angle of Incidence

The angle at which solar radiation hits the leaf (angle of incidence) determines the relative effective radiation intensity on the leaf surface. This angle is defined as the deviation from the sheet normal, where perpendicular and horizontal incident radiation have angles of 0° and 90° respectively. The effective intensity at leaf surface equals the cosine of the angle of incidence (Lambert's Cosine Law).

8 Specifications

Specifications are subject to change without notice.

8.1 LEAF-STATE-ANALYZER LSA-2050

8.1.1 General Design

Housing: Battery-powered handheld device consisting of a control unit and a sample clip, both made of painted polyamide 12 (PA 12). The control unit is equipped with a holder for four AAA-type batteries and a USB-C connector. Two metal flat springs press the clip jaws together. The lower clip jaw is removable.

Display: Backlit transfective B/W LCD display, 48 x 27 mm, 128 x 64 pixel

Control: Six control keys plus a separate START key to initiate a measurement

Data memory: Flash memory, 8 MB, providing memory for more than 30,000 data sets

Data transfer: USB-C port

Power supply: 4 AAA (Micro) rechargeable batteries (eneloop 1.2 V/2 Ah), 4 spare batteries, automatic power/off, battery charger (100 to 240 V AC, 50-60 Hz) for 4 batteries

Operating temperature: -5 to +45 °C, non-condensing

Dimensions: maximum 26.5 cm x 7.0 cm x 3.5 cm (L x W x H)

Weight: 240 g (without batteries)

8.1.2 Measuring Modules

Viewing area: Disk with 10 mm diameter

Upper clip jaw: Five LEDs are circularly arranged around a PIN photodiode, which is shielded from LED emission by a long-pass filter. A quartz glass disk closes the LED/photodiode compartment. Measuring light consists of 10 μs pulses given at 15 Hz except for F_M determinations (100 Hz). Typical maximum emission wavelength, full width at half maximum (FWHM), and integrated intensity at 15 Hz are: UV-B, 310 nm, 15 nm, 0.1 $\mu\text{mol m}^{-2} \text{s}^{-1}$ (0.05 W m^{-2}). UV-A, 365 nm, 12 nm, 0.3 $\mu\text{mol m}^{-2} \text{s}^{-1}$ (0.1 W m^{-2}). Blue, 450 nm, 14 nm, 0.1 $\mu\text{mol m}^{-2} \text{s}^{-1}$. Green, 530 nm, 27 nm, 0.1 $\mu\text{mol m}^{-2} \text{s}^{-1}$. Red, 630 nm, 24 nm, 0.1 $\mu\text{mol m}^{-2} \text{s}^{-1}$. The UV LEDs are only activated in the presence of a fluorescing sample.

Lower clip jaw: A far red LED (peak wavelength 715 nm, FWHM 25 nm) and a near infrared LED (peak wavelength 770 nm, FWHM 30 nm) are positioned in the center of the viewing area. The LEDs are covered by a light-diffusing disk and a quartz disk.

8.1.3 Geospatial Data

A GPS receiver, plus accelerometer, gyro- and magneto-scope sensors add geospatial information to each measurement, including the angle at which sunlight impinges on the leaf. Devices are outlined in Fig. 18, page 52.

8.1.4 Walz Calibration Standards LSA-2050/STD

Fluorescence: Special fluorescence layer mounted on aluminum carrier.

Offset: Non-fluorescent and untransparent black plastic.

Scattering: Special plastic film for scattering of radiation.

8.2 Carrying Case LSA-2050/T

Design: Padded plastic case with handle

Dimensions: 36.0 cm x 30.5 cm x 8.0 cm (L x W x H)

Weight: 920 g

8.3 Battery Charger

Design: Four position intelligent charger for AA or AAA Nickel Metal Hydride (NiMH) or Nickel Cadmium (NiCd) batteries

8.4 Accessory

8.4.1 Darkening Bags LSA-2050/DB

General Design: Set of three small, three medium and three large light-tight bags for dark acclimation of leaves of different sizes made of aluminum foil, colored on the outside. Each bag has a 2 cm diameter central hole for non-invasive determination of chlorophyll concentration. During dark acclimation, both sides of the hole are covered by metallized PET plastic flaps. The flaps magnetically attract each other which ensures that both lie

tightly on the surface. The flaps are flexibly attached to the darkening bag at one end. The other, loose end is folded slightly outwards so that the sample clip of the LSA-2050 can be guided to the hole by folding open the flaps.

Dimensions: 100 mm x 70 mm, 150 mm x 100 mm, 180 mm x 120 mm (small, medium, and large size, respectively).

Weight: 3 g, 4 g, 5 g (small, medium, and large size, respectively)

9 Guarantee

All products supplied by the Heinz Walz GmbH, Germany, are warranted by Heinz Walz GmbH, Germany to be free from defects in material and workmanship for two (2) years from the shipping date (date on invoice).

9.1 Manufacturer's Guarantee

Under this Manufacturer's Guarantee ("Guarantee"), subject to the Conditions and Instructions below, Heinz Walz GmbH, Germany ("Manufacturer"), guarantees (§443 BGB) to the end customer and user ("Customer") that all products supplied by it shall substantially conform in material respects to the Specifications for 24 months from the delivery date (date on invoice). In this Guarantee, "Specifications" means the product's features (as may be amended by Manufacturer from time to time), which are set out under the headings "specifications" and/or "technical specifications" within the product's respective brochure, data sheet, or respective tab on the Manufacturer's website for such product, and which may be included with the documents for the product when delivered. In case of an eligible guarantee claim, this Guarantee entitles the Customer to repair or replacement, at the Manufacturer's option, and this Guarantee does not include any other rights or remedies.

9.2 Conditions

This Guarantee shall not apply to:

- Any defects or damage directly or indirectly caused by or resulting from the use of unauthorized replacement parts and/or service performed by unauthorized personnel.
- Any product supplied by the Heinz Walz GmbH, Germany which has been subjected to misuse, abuse, abnormal use, negligence, alteration or accident.
- Damage caused from improper packaging during shipment or any acts of God.
- Batteries, cables, calibrations, fiberoptics, fuses, gas filters, lamps (halogen, LED), thermocouples, and underwater cables.
- Defects that could reasonably have been detected upon inspection of the product when received by the Customer and not promptly noticed within ten (10) days to Heinz Walz GmbH.
- Submersible parts of the DIVING-PAM or the underwater version of the MONITORING-PAM have been tested to be watertight down to the maximum operating depth indicated in the respective manual. Guarantee shall not apply for diving depths exceeding the maximum operating depth. Further, guarantee shall not apply for damage resulting from improper operation of devices, in particular, the failure to properly seal ports or sockets.

9.3 Instructions

- To obtain guarantee service, please follow the instructions below:
- The Walz Service Information Form available at http://www.walz.com/support/repair_service.html must be completed and returned to Heinz Walz GmbH, Germany.

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- The product must be returned to Heinz Walz GmbH, Germany, within 30 days after Heinz Walz GmbH, Germany has received written notice of the defect. Postage, insurance, and/or shipping costs incurred in returning equipment for guarantee service are at customer expense. Duty and taxes are covered by Walz.
 - All products being returned for guarantee service must be carefully packed and sent freight prepaid.
 - Heinz Walz GmbH, Germany is not responsible or liable for missing components or damage to the unit caused by handling during shipping. All claims or damage should be directed to the shipping carrier.

9.4 Applicable law

- This Guarantee is governed by German law. Place of jurisdiction is Bamberg, Germany.

10 Index

6 mm Diameter Spot for Small Leaves	8	Documentation	38
Accessory	58	Downloading Data	24
Active Tests	18	Epidermis	48
Adjusting Time	24	Excel File	26, 27
Angle of Incidence	54	Extent of Delivery	7
Anthocyanins	45	Flavonoids	46
Battery	7, 56	Fluorescence and Offset Standards	7
Battery Charger	58	FTDI USB Driver	22
Bilger	6	Geodata	17, 50
C4 plant	41	Geodata Hardware	51
Calibration	20	Growth Conditions	39
Calibration and Measurement	49	Guarantee	60
Calibration Data	30	Hydroxycinnamic acids	46
Carotenoids	45	Installing the Software	22
Carrying Case	58	Introduction	5
Cerovic	6	Keys	13
Chl. Correlations	42	Lambert's Cosine Law	54
Chl. Measurements	39	Leaf Slope	53
Chl. Settings	18	LED Combinations	46
Chlorophyll and Small Leaves	44	Local Time	25
Chlorophyll Calibration	38	LSA Download/Time Windows	25
Chlorophyll Determination with Small		LSA-2050 Description	8
Leaves	44	LSA-2050 Info	22
Chromatogram for Pigment		LSA-2050 Measuring Head	9
Determination	40	LSA-2050 Modified for Bulky Samples	10
Connecting the LSA-2050 to Computer	23	LSA-2050 Overview	9
Contents	1	LSA-2050 Software	22
Create New File	21	LSA-2050 Window	15
Darkening Bags LSA-2050/DB	11	Main Menu	17
Detailed Instructions	14	Main Windows	14
Device Info	22	Measure Chart	27
Device Settings	19	Memory	21, 56
		Mesophyll Reference	19

Mesophyll Reference Factor	46	Screening	44
Mesophyll Reference Factors RF	48	Settings	18
N Status Window	15	Sheet Measure	26
Operating Temperature	56	Source Data 01	31
Operation	13	Source Data 02	31
Orient. Sens. Calibration	21	Specifications	56
Plant Species	38	Status Indicators	15
Plant Species used for Chl. Calibration		Sun Position	52
.....	38	Time Setting	25
Quick Start	14	USB Cable	7
References	6	UTC	25
Results	29	UV-A	44
Safety Instructions	3	UV-B	44
SAT Chart	28	Viewing Area	57
Sat Pulse Data	16	Wavelengths Chl Determination	41
Schreiber	6		

11 List of Figures

Fig. 1:	Fluorescence and Offset Standards	7
Fig. 2:	6 mm Diameter Spot for Small Leaves	8
Fig. 3:	LSA-2050 Overview	9
Fig. 4:	LSA-2050 Measuring Head	9
Fig. 5:	LSA-2050 Modified for Bulky Samples	10
Fig. 6:	Darkening Bags LSA-2050/DB	11
Fig. 7:	Keys	13
Fig. 8:	Installation Window	23
Fig. 9:	Access to USB-C Port	23
Fig. 10:	LSA Download/Time Windows	25
Fig. 11:	Measure Chart	27
Fig. 12:	SAT Chart	28
Fig. 13:	Chromatogram for Pigment Determination	41
Fig. 14:	Chlorophyll determination wavelengths	42
Fig. 15:	Chlorophyll Determination with Small Leaves	45
Fig. 16:	Emission spectra of LEDs involved in screening measurements	47
Fig. 17:	Leaf of <i>H. telephium</i> with Epidermis Partially Removed	49
Fig. 18:	Geodata Hardware	52
Fig. 19:	Sun Position	53
Fig. 20:	Leaf Slope	54
Fig. 21:	Leaf Azimuth	54
Fig. 22:	Angle of Incidence	55

12 List of Tables

Table 1:	LSA-2050 Window	15
Table 2:	Status Indicators	15
Table 3:	N Status Window	15
Table 4:	Sat Pulse Data	16
Table 5:	Geodata	17
Table 6:	Main Menu	17
Table 7:	Settings	18
Table 8:	Active Tests	18
Table 9:	Chl. Settings	18
Table 10:	Mesophyll Reference	19
Table 11:	Device Settings	19
Table 12:	Calibration	20
Table 13:	Orient. Sens. Calibration	21
Table 14:	LSA-2050 Info	22
Table 15:	Results	29
Table 16:	Calibration Data	30
Table 17:	Source Data 01	31
Table 18:	Source Data 02	31
Table 19:	Plant Species used for Chl Calibration	39
Table 20:	Growth Conditions	40
Table 21:	Chl. Correlations	43
Table 22:	LED Combinations	47
Table 23:	Mesophyll Reference Factors	49